

Draft Genome Sequences of Ciliavirus and Brinovirus from San Francisco Wastewater

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We report the draft genome sequences of ciliavirus and brinovirus, two members of a likely new family of RNA viruses assembled from San Francisco wastewater. Based on sequence alignments and a nonuniversal genetic code, we believe these to be the first described RNA viruses of ciliates; however, more work is necessary to confirm their host.

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Ciliates are large single-cellular protozoal organisms defined by the presence of cilia as well as macro- and micronucleus. Ciliates are also noted for possessing a nonuniversal genetic code. Virus-like particles have been reported to be associated with *Hyalophysa chattoni*, and a large DNA virus, chlorella virus, has been isolated from *Paramecium bursaria* (1, 2). However, no RNA virus has been recovered from ciliates to date (3).

While performing weekly metagenomic sequencing of San Francisco wastewater, we recovered two contigs of 10,381 and 9,565 nucleotides that distantly aligned by BLASTx to a small portion of an RNA-dependent RNA polymerase of various members of the *Flaviviridae*. Translation of the two contigs using the universal genetic code failed to reveal open-reading frames (ORFs) of >500 nucleotides. However, translation of two contigs utilizing the ciliate genetic code revealed ORFs of 9,954 and >9,260 nucleotides that preserved the BLASTx alignments. HHPred analysis of the contigs translated in the ciliate genetic code revealed a superfamily II helicase similar to eIF4A followed by a picornavirus/ flavivirus-like RNA-dependent RNA polymerase followed by a birnavirus/ sobemovirus-like viral capsid protein, suggesting a genome organization most similar to that of the *Potyviridae* (3, 4). Of note, the two polyproteins were 28.7% identical by amino acid to each other throughout the ORF and 40% identical by amino acid to each other in the putative RNA-dependent RNA polymerase and helicase regions, consistent with these two contigs forming two new genera of a new viral family.

Notably, both contigs were recovered in only one wastewater sample from 25 January 2010 that was taken after a large rainstorm that left >5 inches of rain over the preceding week. Over one-quarter of the alignable nonchordate eukaryotic reads from the DNAsed sample aligned to *Ciliophora* organisms with half of these reads belonging to the *Tetrahymenidae* family, making it one of the most abundant organisms in the sample and strengthening the case for these viruses having ciliate hosts (5).

Sample processing was performed on 1 liter of wastewater that was concentrated to <5 mL with particles between the

sizes of 0.22 μm and 300 kDa using Millipore Pellicon XL 300-kDa filters and 0.22- μm spin columns. Viral particle-enriched sample was treated with micrococcal nuclease and nucleic acid was extracted using a Zymo Viral DNA/RNA kit and half of the recovered nucleic acid was treated with DNase. Both contigs were assembled using PRICE v1.0 and Geneious v8.0 Assembler with metagenomic analysis using SURPI v1.0 from a total of 15,719,690 paired-end 65-bp reads sequenced on an Illumina GAIIX split between these DNAsed and untreated nucleic acid preparations (5, 6). Average coverage of the two contigs was 550 \times and 1271 \times , respectively. Coverage of ciliavirus in the DNAsed sample was 3.5 \times higher than in the untreated sample, while coverage of brinovirus was 2.4 \times higher in the DNase-treated sample, consistent with these being RNA viruses.

Nucleotide sequence accession numbers. The GenBank accession numbers for ciliavirus and brinovirus are [JN661159](https://www.ncbi.nlm.nih.gov/nuclseq/JN661159) and [KF412899](https://www.ncbi.nlm.nih.gov/nuclseq/KF412899), respectively.

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Draft Genome Sequences of Marine RNA Viruses SF-1, SF-2, and SF-3 Recovered from San Francisco Wastewater

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We report the draft genome sequences of marine RNA viruses SF-1, SF-2, and SF-3, which were assembled from metagenomic sequencing of organisms in San Francisco wastewater. These viruses were most closely related to marine RNA virus JP-B and algae viruses.

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The picorna-like superfamily is a rapidly expanding taxonomic unit of positive-stranded RNA viruses with conserved RNA-dependent RNA polymerase (RdRp), capsid, and helicase proteins that have a broad host range, including animals, plants, and insects (1–3). While performing weekly metagenomic sequencing of organisms in San Francisco wastewater, we assembled three contigs of 8,695, 9,270, and 8,642 nucleotides that aligned by BLASTx to the RNA-dependent RNA polymerase and capsid genes of the marine RNA virus JP-B (30 to 40% amino acid identity), a member of the *Picornavirales* order (1, 2). The first two contigs contained bicistronic viral genomes with open reading frames (ORFs) of 4,782/2,877 nucleotides (RNA-dependent RNA polymerase-containing polyprotein ORF/structural polyprotein ORF) and 5,178/2,664 nucleotides, while the third genome contained a single ORF of 7,731 nucleotides in the standard genetic code. The ORFs aligned by 30 to 38% to each other and ~30% by amino acids to RNA viruses of *Chaetoceros tenuissimus*, *Rhizosolenia setigera*, and *Asterionellopsis glacialis*, suggesting that these may be RNA viruses of algae.

All three contigs were assembled from a single metagenomic library derived from a wastewater sample taken on 25 January 2010 following a large rainstorm that left >5 inches of rain over the preceding week. Unlike the likely ciliate viruses also discovered in this sample, wastewater samples collected in March 2010 also contained reads to these viruses but at a significantly lower sequence count. This sample was created by 200-fold concentration of 1 liter of wastewater, with particles between the size of 0.22 μ m and 300 kDa using Millipore Pellicon XL 300-kDa filters and 0.22- μ m spin columns. The viral particle-enriched sample was treated with micrococcal nuclease, nucleic acid was extracted using the Zymo viral DNA/RNA kit, and half of the recovered nucleic acid was treated with DNase. The three contigs were discovered and assembled using PRICE version 1.0 (4), the Geneious version 8.0 Assembler, and SURPI version 1.0 (5) from a total of

15,719,690 paired-end 65-bp reads sequenced on an Illumina GAIIX split between these DNAsed and untreated nucleic acid preparations (3, 6). The average coverages of the three contigs using all reads from the sample were 1,749 \times , 1,379 \times , and 222 \times .

Nucleotide sequence accession numbers. The GenBank accession numbers for marine RNA viruses SF-1, SF-2, and SF-3 are JN661160, KF412901, and KF478836, respectively.

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Draft Genome Sequences of *Leviviridae* RNA Phages EC and MB Recovered from San Francisco Wastewater

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We report here the draft genome sequences of marine RNA phages EC and MB assembled from metagenomic sequencing of organisms in San Francisco wastewater. These phages showed moderate translated amino acid identity to other enterobacteria phages and appear to constitute novel members of the *Leviviridae* family.

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Leviviridae is a family of positive-stranded RNA bacteriophages consisting of two genera, *Allolevivirus* and *Levivirus*, which are each generally restricted to a single host genus of Gram-negative bacteria (1). Of note, the RNA-binding coat protein of the *Levivirus* MS2 phage has served as a tool for many basic science applications and as a model for the assembly of RNA viruses (2, 3). Interestingly, surveys performed in 1976 and 2006 of marine/coastal RNA virus communities found very few if any RNA phages (4).

While performing weekly metagenomic sequencing of organisms in San Francisco wastewater, we assembled 3,180-nucleotide (EC) and 3,925-nucleotide (MB) contigs at 849× and 229× coverage, respectively, which aligned by BLASTx to enterobacterial phages Hgal1, M11, and Qbeta. Each contig comprised a typical *Leviviridae* genome organization, consisting of three open reading frames (ORFs) encoding maturation, coat, and replicase proteins, with phage EC likely truncated by ~600 nucleotides at the 3' end. However, neither phage encoded a readily apparent lysis or read-through protein. Phage EC appears to possess ORFs that encode both coat and replicase proteins but lacks any protein with alignment to known lysis proteins of *Levivirus*. Phage MB also appears to lack a recognizable lysis protein.

The predicted replicase protein of phage MB demonstrated 39% amino acid identity to *Caulobacter* phage phiCb5 over the entirety of the replicase protein, while the closest replicase to phage EC demonstrated 42% amino acid identity to enterobacterial phage Hgal1. Both replicases demonstrated <38% identity to members of *Leviviridae* already placed in a genus, which is equivalent to amino acid identity between *Allolevivirus* and *Levivirus* genera, consistent with these two viruses forming two novel genera within *Leviviridae*. The predicted maturation proteins aligned with 32% amino acid identity to enterobacterial phage BZ13 (EC) and 28% to *Caulobacter* phage phiCb5 (MB). The predicted phage EC coat protein aligned with 32% amino acid identity to enterobacterial phage C-1 INW-2012, while the predicted phage MB coat protein failed to demonstrate significant amino acid alignments to any phage proteins but did significantly align by HHPred to *Caulobacter* phiCb5 virus-like particle (5, 6).

The sample that yielded these draft genome sequences was prepared from 1 liter of wastewater that was concentrated to <5 ml of particles between the size of 0.22 μm and 300 kDa using Millipore Pellicon XL 300-kDa filters and 0.22-μm spin columns. Nucleic acid was extracted using the Zymo viral DNA/RNA kit, and half of the recovered nucleic acid was treated with DNase. The phage genomes were discovered and assembled using PRICE version 1.0, Geneious version 8.0 Assembler, and SURPI version 1.0 from a total of 15,719,690 paired-end 65-bp reads sequenced on an Illumina GAIIx split between these DNased and untreated nucleic acid preparations (7, 8).

Nucleotide sequence accession numbers. The GenBank accession numbers for marine RNA phages EC and MB are [KF616862](https://www.ncbi.nlm.nih.gov/nuclot/KF616862) and [KF510034](https://www.ncbi.nlm.nih.gov/nuclot/KF510034), respectively.

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