

Array Washing Protocol

-It is recommended that all wash stock solutions be filtered before using.

- 1. Prepare wash solutions in glass slide dishes, with each dish having its own rack.**

**Wash Solution I: 340 ml Milli-Q water
 10 ml 20XSSC
 1 ml of 10%SDS**

**Wash Solution II: 350 ml Milli-Q water
 1 ml 20XSSC**

- 2. Carefully remove array from water bath, making sure to keep chamber level. Dry hyb chamber with paper towels and attempt to “wick” any water away from chamber seams.**
- 3. Unscrew chamber and remove array. Some water may enter chamber and pool under slide at this time. If so, it is helpful to have a pair of forceps to pry array away from chamber.**
- 4. Keep array level when submerging in Wash I. Once submerged, tilt array and gently dump off coverslip. It may be necessary to lightly swish array under solution to dislodge the slip.**
- 5. Once slip is off and laying on bottom of slide dish, put array in rack and remove any additional hybs from water bath. When all chips are in Wash I, plunge rack up and down 10-20 times.**
- 6. Individually transfer chips to slide dish containing Wash II, do not transfer entire slide rack as this will cause too much SDS carryover.**
- 7. Dry array in room temperature table top at 600 rpm for 5 min.**
- 8. Try to scan array within hours of washing as the Cy dyes are unstable and will degrade differentially.**