Protocol for Precipitation and 384 Well Arraying of PCR Products

1. After running 96 well reactions on 1% agarose gel and documenting results, add 10µl Sodium Acetate, pH 5.5, and 110µl room temperature Isopropanol.

2. Transfer reactions to a Costar 96-well U-bottom Polypropylene Plate (#3790). To save on pipette tips, first add IPA to Costar plates and then transfer PCR + NaOAc pH 5.5 to IPA. Let plates precipitate at -20° C for at least overnight. This transfer to new plate allows centrifugation of 18 plates simultaneously in an RC3B centrifuge using 6 plate carrier adapters. To spin, plates must be taped together correctly. Please see experienced users for a demonstration.

3. Spin plates at 4500 rpm for at least 2 hours.

4. Carefully aspirate solution using a 12 channel Wheaton Aspiration Adapter from PGC Scientific, 800-424-3300, catalog #851388.

5. Add 100µl of 70% EtOH. Spin plates for another hour at 4500.

6. Aspirate again and let air dry or dry in 96 well speed-vac.

7. Allow PCR products to resuspend in 20µl of H2O for at least 18 hours.

8. Transfer products to Genetix 384-well printing plates (available from USA/Scientific Plastics, 800-872-3847, product #2802-0384).

9. Dry plates down in speed-vac, resuspend products in an appropriate volume of 3X SSC.

10. Let plates resuspend at least overnight before printing.

For additional information about various precipitation options, see:
http://cmgm.stanford.edu/pbrown/protocols/ASH/pcrcleanupcomparison.htm